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Managing dose-, damage- and data-rates in multi-frame spectrum-imaging

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Abstract

As an instrument, the scanning transmission electron microscope (STEM) is unique in being able to simultaneously explore both local structural and chemical variations in materials at the atomic scale. This is made possible as both types of data are acquired serially, originating simultaneously from sample interactions with a sharply focused electron probe. Unfortunately, such scanned data can be distorted by environmental factors, though recently fast-scanned multi-frame imaging approaches have been shown to mitigate these effects. Here, we demonstrate the same approach but optimised for spectroscopic data; we offer some perspectives on the new potential of multi-frame spectrum-imaging (MFSI) and show how dose-sharing approaches can reduce sample damage, improve crystallographic fidelity, increase data signal-to-noise (SNR), or maximise usable field of view. Further, we discuss the potential issue of excessive data-rates in MFSI, and demonstrate a file-compression approach to significantly reduce data storage and transmission burdens.

Keywords:

Spectrum imaging, non-rigid registration, beam damage, dose-rate, data compression

Introduction

The aberration corrected scanning transmission electron microscope (STEM) can now routinely image crystalline specimens at atomic resolution. Of the imaging modes available the relative ease of interpretation of the annular dark-field mode (ADF) makes this a popular choice, offering thickness / atomic-number type contrast. However, the mixed thickness-composition contrast introduces some ambiguity in heteronuclear samples, so for composition studies either energy dispersive x-ray analysis (EDX) or electron energy-loss spectroscopy (EELS) are added to many STEMs. Unfortunately, the collection efficiency of these signals is around 100 and 10,000 times weaker respectively compared with imaging [1].

While spectrometer hardware is improving gradually, little has changed in the way that spectrum images (SI) are recorded since their introduction [2,3]. For most studies, the poor collection efficiency means the signal-to-noise ratio (SNR) limits the clarity of results. There are three common approaches pursued to increase the SNR of elemental maps; increasing the beam current, increasing the pixel dwell-time or installation of a higher collection efficiency detector or spectrometer. However, the first two approaches have significant drawbacks. Increasing beam current can rapidly cause sample damage [4] or accelerate carbon contamination, while scanning slower risks introducing scanning distortions, stage-drift or focal-drift into the frame [5]. Further, it is not always possible to increase beam current (such as in monochromated systems), while increased dwell-times make recording larger fields of

view prohibitively time consuming for the operator. Lastly, spectrometer upgrades may not be possible, for example due to objective pole-piece space restrictions or prohibitive cost.

Thanks to recent developments in the robust registration of serial scanned image data [6,7], multi-frame spectrum-image (MFSI) data can now be equally successfully reregistered [8,9]. Multi-frame experiment design frees the operator from using slow scans and high-probe currents to realise acceptable SNRs. Instead, the required signal is built up from a series of faster and individually lower dose scans; specimen drift is greatly reduced, as is in many cases beam-damage, while the redundant set of complimentary ADF images allows for scanning distortion to be compensated without the need for specially designed instruments [10].

Since the term ‘spectrum imaging’ was introduced in the 1980s, continuous improvements in instrumentation and computer power have been realised. However, MFSI inevitably generates a far greater data-rate than conventional single scan approaches. Approaches to reduce EELS-SI data-rate based on compressed-sensing have been proposed, but these require additional hardware such as grating apertures and complex synchronisation [11]. As data-rates increase, there is wisdom in returning to those suggestions made before computing power was so advanced. Previous approaches have considered data-binning [2], or neighbour-difference approaches [12]; however, these may be unacceptably lossy or offer only limited storage savings. Here we seek to implement some approach which requires no new hardware and instead makes use of the prior knowledge about the regions of the spectrum which contain more information than others, and about how experimentalists ultimately process the data (with background fitting and integration) to produce various 2D maps. We present a modification of the method proposed in [2], but dividing SI data in the

energy domain into blocks, some of which receive no compression where fine measurements are needed (such as the zero-loss peak (ZLP) position or fine-structure edges), and others which are down-sampled after some appropriate noise reduction.

In summary, in this work we make use of the ability to acquire, store, and robustly align MFSI data to introduce a new free-parameter into the operator's experiment design. We present five separate studies, each highlighting different aspects of this new approach that probe the performance of the method. In this work we:

1. present a fixed-electron-dose study, comparing the sample-damage of a beam-sensitive specimen with and without multi-frame dose-fractionation,
2. demonstrate (for a beam-tolerant specimen) the accumulation of SNR, and the utility of digital super-resolution in spectrum imaging,
3. show a sub-20pA atomic-resolution EDX mapping example, through the combined use of MFSI alignment and image template-matching (TM),
4. reveal through the use of multivariate statistical analysis (MSA), the minimum dose required to capture the information-content of SI data, and
5. describe a new highly-efficient data-compression routine to minimise the data-management burden of recording multi-frame EDX SI series.

Methods

In this work, several different descriptors of sample-beam exposure are used; for clarity, these are defined explicitly here. *Dose-per-frame* is expressed in units of electrons per-square-Angstrom ($e^- \text{\AA}^{-2}$). The *total-dose*, i.e. the entire electron-dose, is also expressed in units

of electrons per-square-Angstrom ($\text{e}^{-}\text{\AA}^{-2}$) and is simply the dose-per-frame multiplied by the number of frames in the series. *Dose-rate* is the total-dose divided by the duration of the corresponding exposure, with units of electrons per-square-Angstrom per-second ($\text{e}^{-}\text{\AA}^{-2}\text{s}^{-1}$); this is either calculated as the dose-per-frame divided by the frame-time (number of frame pixels multiplied by the dwell-time), or as the total-dose divided by the total series-time. A final commodity which is of merit for fast-scan multi-frame acquisitions where the probe is always moving, is the instantaneous pixel charge delivered by the passing probe (in electrons), and is calculated as the beam current multiplied by a single pixel dwell-time.

Samples and Hardware Configurations for each Test

Three unique instruments were used across the studies presented here. For the fixed-dose study (test 1), the sample used was $\text{Pb}_2\text{ScTaO}_6$ prepared for STEM imaging by ion beam milling. This material exhibits incomplete long-range ordering on the Sc:Ta sublattice. Two data-sets were recorded using a doubly-corrected JEOL ARM200F with a ($\approx 1\text{\AA}$ probe of 74pA with a convergence of 30mrad). Simultaneous hardware synced spectra were recorded using a Gatan Quantum GIF and Oxford Instruments X-max 80mm² detector. A single 256x256 probe-position spectrum image with a dwell-time of 0.01 s/pix was recorded, and for comparison five separate spectrum images with a dwell-time one fifth of the single frame (0.002s/pix), i.e. equal total-electron-dose and equal dose-rate. Imaging was carried out near an anti-phase boundary.

To demonstrate SNR accumulation, digital super-resolution, and EELS SI data-compression (test 2 & test 5), an A-site deficient perovskite, $\text{Nd}_{0.6}\text{Ca}_{0.1}\square_{0.3}\text{TiO}_3$ (where \square denotes A-site vacancies), was used. The sample was prepared by conventional crushing (pestle and mortar) and drop cast on a standard 3mm lacy-C film support; the sample was

previously characterised and discussed in [13]. ADF imaging and atomically resolved EELS mapping were performed using a NION UltraSTEM100 MC dedicated STEM equipped with a Gatan Enfium ERS spectrometer optimised with high stability electronics. The microscope was operated at 60kV with the probe forming optics configured to form a $\approx 1\text{\AA}$ probe (full-width at half-maximum), with a convergence semi-angle of 31mrad yielding a probe current of $\approx 110\text{pA}$. The dataset presented here comprises of eight spectrum images with lateral dimensions of 60x94 pixels ($\approx 0.67\text{\AA}$ pixel-size) acquired consecutively using a dwell time of 0.001s/pix and 16 x 16 subpixel scanning, into a spectrometer entrance aperture with a 48mrad semi-angle. The total acquisition time per SI frame was 1min 15s, including overheads. No drift compensation was applied during the acquisition or between SI frames.

For the MFSI, TM, and data compression (tests 3-5), an $\text{Ag}_2\text{ZnSnSe}_4$ (AZTS) sample was prepared from a stoichiometric mix of pure elements annealed at up to 510°C for 46 days in sealed ampoules at 10^{-6} torr, then slow-cooled at 3°C/hour to room temperature achieving grain sizes of up to several micrometers. One extended single-crystal was imaged which at the edge was determined by EELS t/λ to be $\approx 20\text{-}30\text{nm}$ thick but extended laterally for several hundred nanometers. Annular dark-field (ADF) imaging and EDX mapping was performed using a JEOL ARM200CF equipped with a single Centurio 100mm² EDX detector. A 17pA beam at 200kV was used to record 230 simultaneous ADF-EDX SI pairs at 200x200pixels ($\approx 1\text{\AA}$ probe, $\approx 0.56\text{\AA}$ pixel-size) at a rate of 1000spectra/sec. Frame time was approximately 41s. This gives a dose-per-frame of $3.44 \times 10^5 \text{ e}^-\text{\AA}^2$. Images across the single crystal were recorded in sets of 10-frames, each set from a virgin area of crystal.

Data Acquisition

To reduce operator burden and maximise experimental throughput, an interactive graphical script was written in Digital Micrograph where image-SI pairs are recorded, numbered, and saved to disk automatically. Figure 1 shows the appearance of the developed user interface.

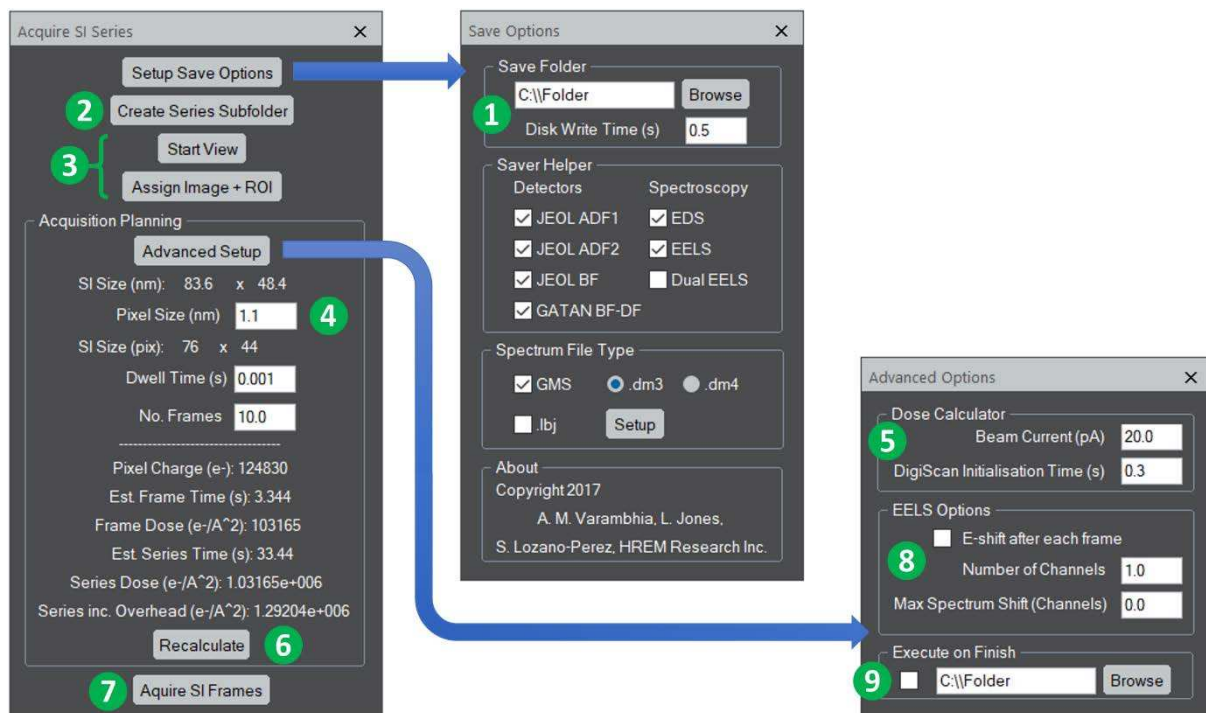


Figure 1. Interactive graphical user interface for the streamlined data acquisition, numbering and saving. The numbered items are described in the main text.

Here the user first indicates the save-path for the microscope session (1), the data writing overhead time, the image and SI signals to record, and for the SI whether file-compression should be used; after that numbered sub-folders may be created for each new region-of-interest (ROI) without leaving the main work-flow (2). After this the user works down; to start a live image-view, capture the survey image, and specify the region of interest (3). With the position and size of the ROI set, the user specifies the required pixel-size (4); this is contrary to the microscope's built-in workflow where the number of pixels is treated as the free parameter. Here the number of pixels is automatically determined for the user to deliver them a specific pixel size, with a minute expansion of the ROI size if necessary.

In the 'Advanced Setup' menu, the user specifies the STEM probe current and the time-overhead of initialising communications to the DigiScan unit (5).

With the SI parameters set, the user is presented with estimates of the frame-time, series-time and their associated electron-dose estimates. If the user adjusts either the size of the ROI, the pixel-size, dwell-time or number of frames, the time and dose estimates can be re-evaluated (6). This gives the user an indication of the practicability of their set conditions *before* the acquisition is started. Detailed spectrometer settings such as EELS dispersion, offsets and exposure times are handled as usual in the Gatan menus. The electron-dose for the entire series including all communication and fly-back overhead times is also shown; this is often ignored but may be crucial for studies on beam sensitive materials.

The image and SI series is then recorded (7) with each data being saved to disk and closed from memory as it is completed. If the sample remains undamaged, and further SI frames are desired (for example, to extend the series to increase SNR), the 'acquire SI frames' button can simply be pressed again and further SI frames will be added to the same sub-folder. This can be repeated as required until the user presses the button to create and move to the next numbered sub-folder for the session. In advanced settings, the user can select other useful automations; such as to step the EELS spectrum along the camera between SI frames by incrementing the drift-tube voltage (8) (similar approaches have been used by Bosman & Keast with so-called 'binned gain averaging' [14], or by Hou [15] and later by Wang et al [16] to improve dark-reference/spectrum quality), or to execute a custom script on completion (9). Such a custom script might for example: drop the magnification 10,000x (reduces dose-rate by 10^8), close the beam entirely (on systems where this is available), or to send an email to the operator that the series is complete [17].

Data Processing

Using the ADF images acquired alongside the SI data, the image-alignment and scan-distortion corrections were diagnosed in post-processing following the method in [6]. This approach allows the operator to choose to record scan frames with larger fields-of-view than would otherwise be possible, increasing the accessible field-of-view and reducing the need to chase features by manual adjustment during acquisition [18]. The shift-vectors *between* frames, and the non-linear 2D distortion-fields *within* each frame are stored at the end of this calculation. These data are then used to correct for the equivalent shifts and distortions in the SI data conceptually similar to the method followed by Yankovich et al. [9], but instead utilising specific prior knowledge that the STEM is a scanned instrument, to improve the alignment speed, robustness and accuracy [6]. Moreover, the method deployed here differs from this earlier work, in that the spectrum-image rearrangement and restoration steps (steps 3 and 5 in Figure 3 of reference [9]) are avoided to improve calculation speed and memory use. In the present work, each SI is operated on in series; where within each given SI, every spectral-channel is corrected by the distortion-field for that scan, before being added to a running summation SI-volume. This approach means that at no time do the SI data need to simultaneously stored in computer RAM, all memory needed can be pre-allocated at the start of the calculation, and thus the calculation scales more favourably with both the field-of-view and the number of frames in the SI acquisition. The significance of this serial-SI processing will be discussed further in the context of data-compression later on.

To perform the template-matching, where repeating features are identified across the view [19], a region-of-interest (ROI) is first drawn to highlight a motif within the image. In this case, the selected ROI is 2-3 unit-cells in size and is then cross-correlated across the full field-

of-view to identify all the repeat units of that motif. These repeats are then themselves cropped to form an image series for averaging. Where a corresponding SI exists, the same regions from this can also be cropped and averaged further improving SNR [8,20]. Where an image and SI *series* exist, the image motif can be automatically searched throughout the entire image series (folder), noting the frame number and x-y position before repeating the same across the SI series.

For test 4, the inspection of data information-content by multivariate statistical analysis (MSA), Scree plots were calculated using the Hyperspy package after singular value decomposition (SVD) [21], for increasing cumulative doses. With these, the percentage contribution to the data variance is plotted, where the number of components with a variance contribution visibly greater than the Scree-plot background is an indication of the ease of separability between genuine signal and random noise [22].

For test 5, to compress the EDX file-size, only non-zero data are stored ($\approx 0.04\%$ of SI voxels) noting their position in voxel order and their (integer) x-ray photon count. This data is read and restored to the full dimensions on decompression. To reduce EELS spectra file-size, the over-redundant nature of each spectrum is exploited. First, the energy-domain is divided into regions by the user which are to be used for broad background determination rather than fine feature extraction. While this step requires prior knowledge of the spectral regions of interest, these are often known from say an earlier point spectrum before any mapping is performed. Next, within only the spectral-blocks which are to be down-sampled, extreme outliers are removed from the data (by say median filtering) and random uncorrelated noise (by principal component analysis (PCA) but with several tens of components preserved), before being smoothed in the energy dimension only by a kernel whose width in channels is

equal to the down-sampling. These de-noised data are then down-sampled, and the reduced size SI data are stored *after* taking their natural logarithm. To further reduce storage size, the data are rounded to their first four significant figures. To decompress the data back to the full dimensionality, the missing values are linearly interpolated, before restoring the magnitude by raising them by the natural exponent. This linear interpolation *before* raising by the exponent exploits the power-law decay often found in background regions for thin samples (with no multiple scattering). This is an improvement over the simple binning proposed in [2], and prevents long flat, or even linear blocks, being restored in the decompressed spectra. Our proposed compression is customisable, is locally lossless in the regions where no down-sampling is performed (for example for fine structure studies, or ZLP drift tracking), and allows for significant data-savings in less important regions of the spectrum. The only additional data to store during the compression is the channel-number/down-sampling pairs as annotated in Figure 8.

To produce elemental maps, power-law backgrounds were fitted before the EELS edges, and for the EDX maps characteristic x-rays were integrated after subtracting a linear background. In all comparisons, the same backgrounds and peak/edge integrations were used on both data.

Results

The results presented follow the five themes listed above.

1 – Reduced Damage with Fast Scanning

Figure 2 shows the fixed-dose comparison between conventional and MFSI approaches. As the beam current remained fixed for this test, and as 5-frames at 5x speed

were recorded, the dose-rate ($1.03 \times 10 \text{ e}^{-}\text{\AA}^{-2}\text{s}^{-1}$) and total-dose ($6.7 \times 10^7 \text{ e}^{-}\text{\AA}^{-2}$) were necessarily equal.

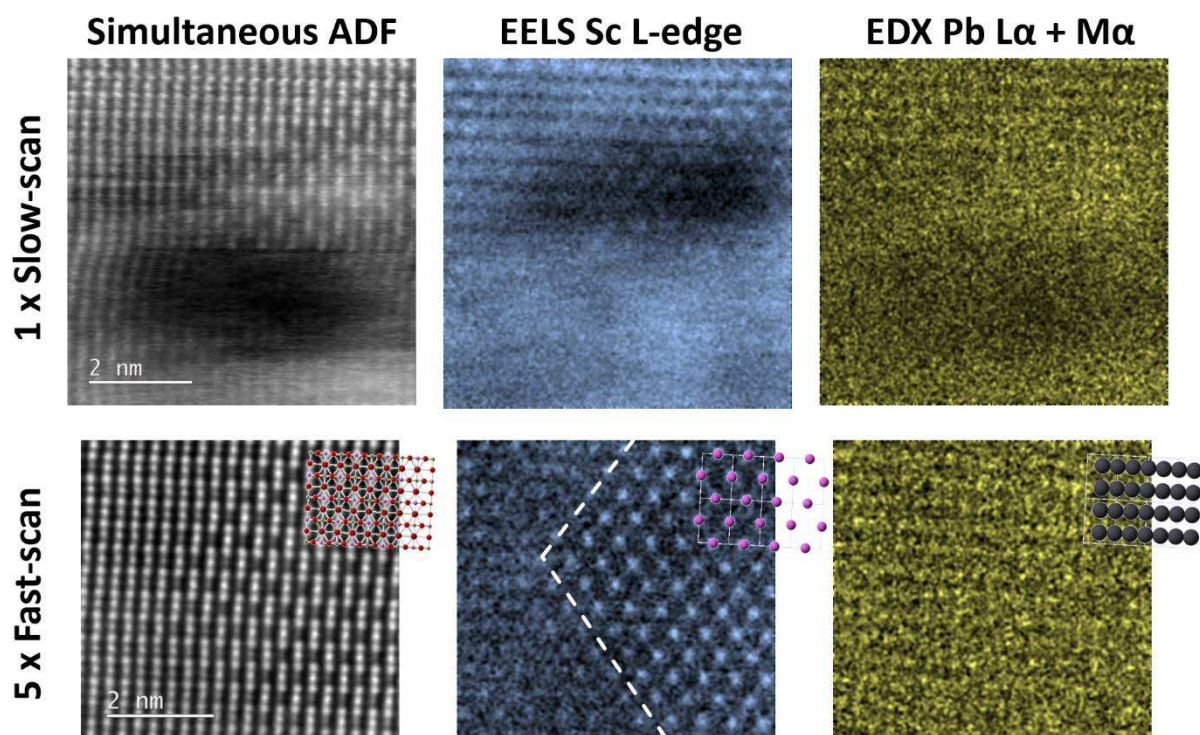


Figure 2. Fixed total-dose comparison of single-scan and multi-frame strategies. Figure shows the ADF image (left), the Sc map measured by EELS (centre), and the Pb EDX map (right). The dashed-line indicates the position of the anti-phase boundary.

The ADF of the conventional single frame acquisition immediately shows the beam-damage and scanning-distortions which arise from using the slower scan speed. The fast-scanned, distortion-corrected, and averaged ADF image however shows both improved SNR and less distorted lattice planes as expected [23]. The location of the anti-phase boundary is now clearly resolved (where the periodicity in the Sc map changes). The SNR of the Sc $L_{3,2}$ -edge EELS maps show an improvement from 2.1 to 5.1 with the Sc positions relating directly with the ADF image. EDX maps are less clear but in the multi-frame average spectrum image the Pb lattice begins to become apparent. As sample damage is reduced so drastically, more frames could have been recorded to improve SNR further if desired. This was not done here to allow a fair comparison at fixed total dose conditions.

This test indicates that in addition to discussing a sample's critical dose, or dose-rate [24], we should perhaps also evaluate the instantaneous pixel-charge, as it is only the third of these quantities which is different in the fast MFSI acquisition. For this example, the pixel charge was reduced from $4.62 \times 10^6 \text{ e}^-$ to $9.24 \times 10^5 \text{ e}^-$ by changing to the MFSI regime.

2 – Improving SNR, Detectability & Sampling (Digital Super-resolution)

Figure 3 shows both a single ADF and EELS-SI frame (top row) displayed with the as-acquired pixel sampling. While the raw data were acquired well above the Nyquist frequency capturing all the resolvable information ($\approx 0.67\text{\AA}$ step size with $\approx 1\text{\AA}$ probe), the resulting maps appear rather pixelated. Instead of the temptation of increasing the sampling during the acquisition (with the associated dose and frame-speed implications), we employ digital super-resolution during the post-processing [25]. The image and SI data were up-sampled by a factor of 4 *before* rigid and non-rigid alignment [6]. It is essential that this up-sampling is performed before alignment for any digital super-resolution benefit to be realised. The bottom row then shows the cumulative ADF image and EELS maps from 8-frames of acquisition.

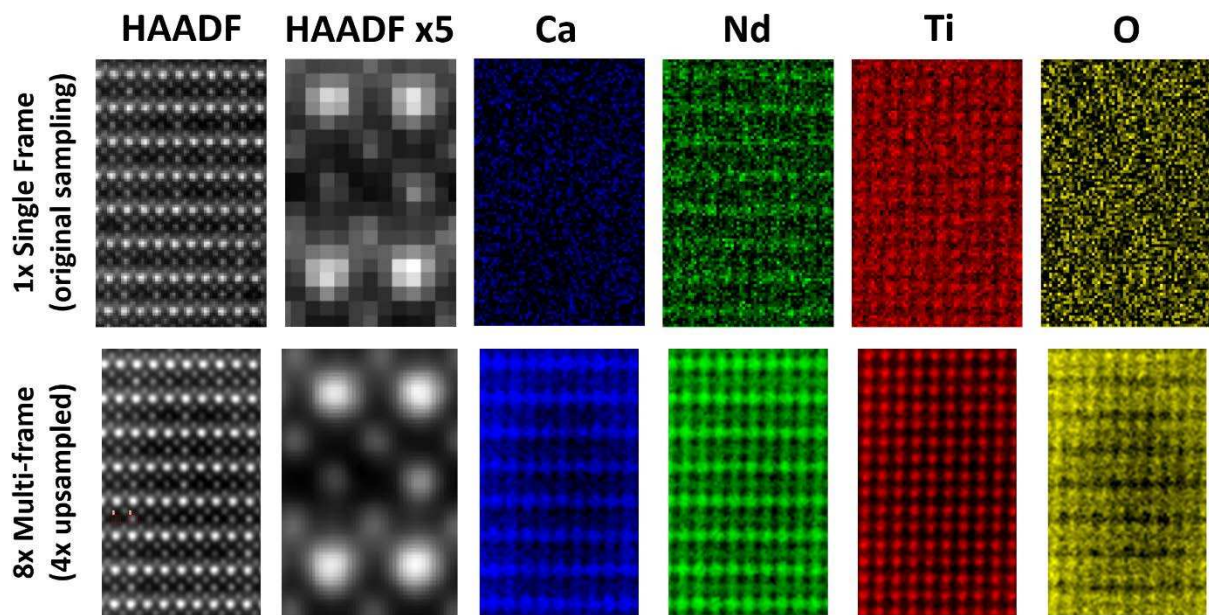


Figure 3. Example of combined use of multi-frame SNR accumulation, and digital super-resolution to increase pixel density.

Image / SI field of view = 40 x 63nm

While some lattice information is visible in the single-frame EELS maps for the Nd and Ti signals, the SNR is very poor; Ca and O maps reveal no periodicity. Upon averaging all four signals are clearly resolved. As in previous studies full intermixing between the Ca and Nd was confirmed. It should be noted that there is some small loss of image-width as a result of cropping following the image alignment (as a result of stage drift), as well as some early signs of damage beginning to appear, especially on the oxygen lattice. This damage may have begun earlier during the operator refocussing and stigmation just before the start of the acquisition whose cumulative effects should be considered in future low-dose studies.

3 – Combining MFSI and Template Matching for sub-20pA EDX Mapping

Image template-matching (TM) with crystallographic prior knowledge has been shown to improve image quality [19]; and further, when exploiting rotational-symmetry has previously been shown to yield atomic resolution EDX maps from within aluminium alloy precipitates [8]. In the example here, during both the acquisition and the analysis, only translational (and not rotational) symmetry was assumed.

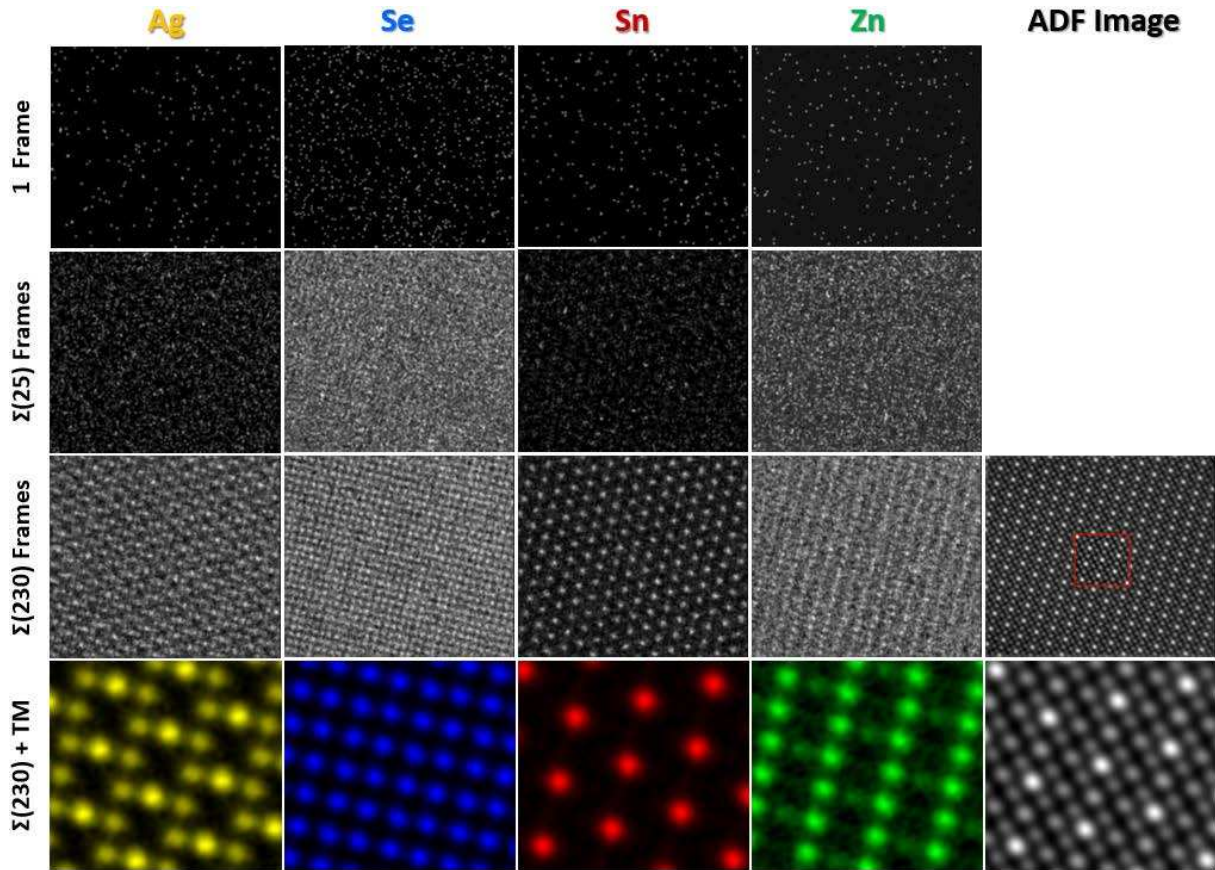


Figure 4. Signal accumulation over a 230-frame EDX series. Far right column shows the aligned ADF image corresponding to the 230-frame series and (highlighted in red) the ROI used for the template-matching (the motif).

Figure 4 shows extracted maps corresponding to the four elements present for a single-scan, after aligning 25 frames, and after summing across the full 230-frame acquisition. These correspond to total-doses of 3.4×10^5 , 8.6×10^6 , and $7.9 \times 10^7 \text{ e}^- \text{Å}^{-2}$ respectively.

After this the ADF ROI shown far bottom-right was used to identify 112 repeat motifs for further SNR accumulation (Figure 4 – bottom row). The effective equivalent dose then of the TM EDX maps is 230x112 times the original single frame dose at approximately $8.8 \times 10^9 \text{ e}^- \text{Å}^{-2}$. This template matching delivers (in post processing) a two-dimensional equivalent of the previously proposed 1D smart line-scans [26], where dose is shared across equivalent positions. In these template-matched EDX

maps, the atomic columns are now clearly resolved and differences in the occupancies are readily visible in the Ag and Zn maps; all the while using only a 17pA STEM probe. Using TM to deliver such high SNR, it may be possible to determine interface intermixing profiles more accurately without needing to increase sample-thickness or beam current [27], or to study the x-ray emission from individual x-ray lines separately [28]. Moreover, TM performed in real-space opens the possibility to map point defects/dopants that would be impossible by Fourier based techniques.

4 – Quantitative Evaluation of Information Content

In this section we continue the analysis of the experimental data from the sample used in section 3 – to determine the *information content* of the recorded spectra. When decomposing spectra using PCA, a Scree plot is used to show the contribution to the variance described in each component. In a decomposition of n-spectra we expect n-components, though in practice far fewer of these orthogonal components contain useful information – this is the basis of the noise reduction. In an ideal situation, the decomposition results would show that close to 100% of the variance is described in as fewer number of components as the sample complexity requires.

Figure 5 shows the Scree-plot data as a function of increasing cumulative electron dose which the sample received. At the lowest dose (single-frame, $3.4 \times 10^5 \text{ e}^- \text{Å}^{-2}$), information is shared almost equally amongst all components, which contain mostly noise. Only for doses greater than $10^6 \text{ e}^- \text{Å}^{-2}$ the first component starts to contain statistically relevant information and a cut-off for denoising can be determined. Further data points represent the sum over the first 1, 4, 9, 16, 25, 36, 49, 64, 81, 100, 121, 144, 169, 196, and 230 frames [21]. A scree plot was also calculated for the template matched summation.

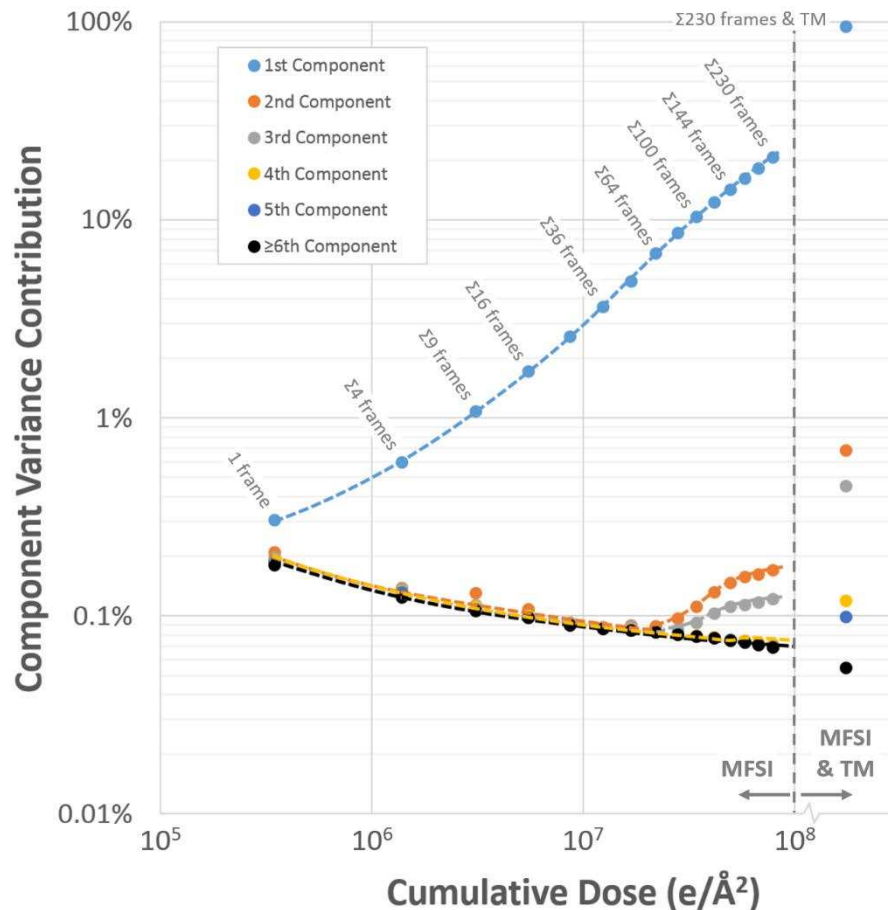


Figure 5. Information content, as evaluated by the component variance in the first 5 components, as a function of increasing dose. Components 6 and onwards represent the noise floor. The increasing dose-series is created by integrating increasing numbers of scan frames as labelled. The sum over the whole 230-frames with TM yields a final data-point (far right) with an effective dose of $8.9 \times 10^9 \text{ e} \cdot \text{\AA}^{-2}$.

As the dose is increased (by summing more frames) the difference between this first and other components becomes greater. From around $2.2 \times 10^7 \text{ e} \cdot \text{\AA}^{-2}$ (64 frames) the second component becomes significant above the noise, then at $2.8 \times 10^7 \text{ e} \cdot \text{\AA}^{-2}$ (81 frames) the third, and at $5.8 \times 10^7 \text{ e} \cdot \text{\AA}^{-2}$ (169 frames) the fourth. After template matching, 5 components were clearly identifiable above the noise (6th onwards).

5 – Managing Data-rates with Real-time Data-compression

While dose-fractionation across n-frames was shown above to improve spatial fidelity and reduce sample damage, it inevitably requires an n-fold increase in computer storage. This situation is especially acute for the example given in Figure 4 where 230 EDX-SI and ADF-image pairs were recorded. The total storage used for this *one region of interest* was 70.3Gb

(75.6×10^9 bytes). This becomes a significant burden both on EM-facility servers but also in transmitting data to collaborators or to analysis PCs and for data-archival purposes.

The solution here lies in the nature of EDX SI-data itself. At low electron-doses, these files are mostly filled with zeros, while (because of the photon counting nature of the EDX detector) the non-zero elements are filled with integer counts. Instead of storing the full 3D arrays, massive storage savings can be realised by listing only the bins that contain non-zero elements. Here we introduce such a *listed-bin-journal* file type (*.lbj) which records data in this sparse way. Figure 6 shows the first frame of the data from Figure 4 compressed from both .dm3 > .zip, and from .dm3 > .lbj.




Name	Type	Size
 EDX_Test_Spectrum.dm3	Gatan DigitalMicrograph Image...	320,626 KB
 EDX_Test_Spectrum.zip	Compressed (zipped) Folder	417 KB
 EDX_Test_Spectrum.lbj	LBJ File	83 KB

Figure 6. Example of the original and compressed EDX SI files as seen in Windows File Explorer. The compression ratio of the .lbj file is approx. 3900x and is a further 5x smaller than a simple .zip file.

The .lbj file is 3,905 times smaller which more than compensates for the acquisition strategy of fractionating the dose across 230 frames. The result is also more than 5x smaller than simply storing the data in a .zip archive. Unlike the >70GB raw data for this dataset, the entire 230-frame EDX series after compression requires ~22MB and can, for example, be sent to a colleague as an email attachment.

To benchmark the performance of the compression algorithm, the same process was repeated for many SI example spectra from a variety of samples and beam currents, yielding varying degrees of SI sparsity.

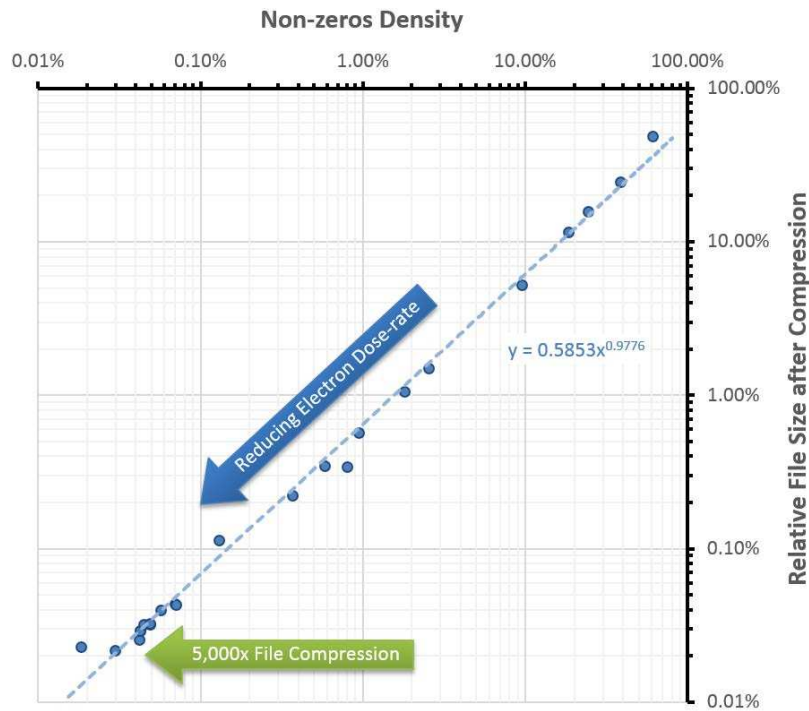


Figure 7. File compression results for increasingly low electron-dose EDX SI volumes.

Figure 7 shows that as the density of non-zero SI elements falls, the compression approach followed here becomes more efficient as expected. For data with very few non-zero elements the files are still compressed but less so. In the tests compressions of up to 5,000x were realized for the lowest dose data.

To evaluate the EELS data compression described in the Methods section, the MFSI data presented in Figure 3 was compressed using blocks of down-sampling ranging from 1 at the titanium edge (no down-sampling), to 60 in the extended background between edges.

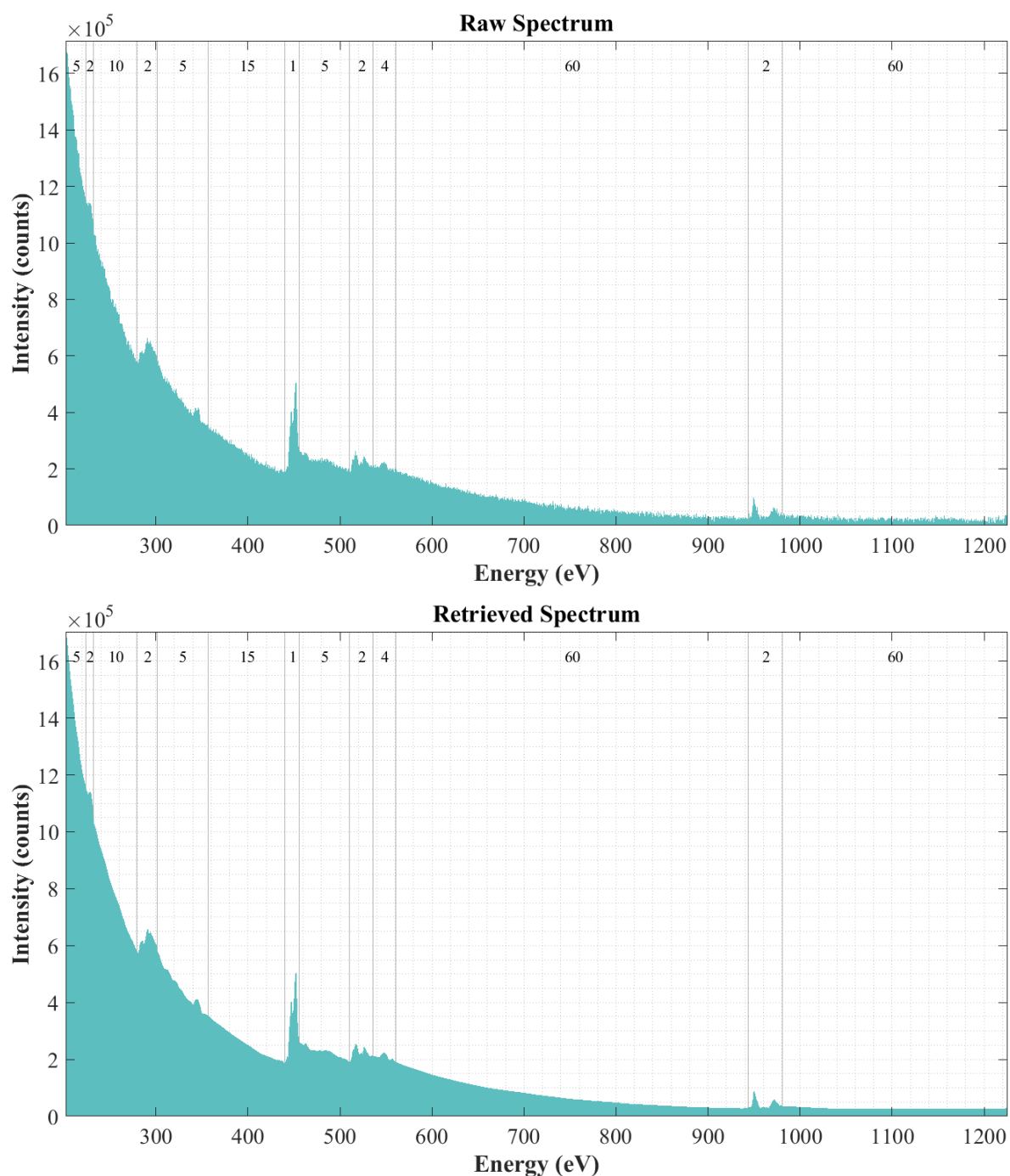


Figure 8. Original EELS spectrum (top) corresponding to one SI of the data shown in Figure 3, and the equivalent spectrum after compression and decompression (bottom). Annotations across the top of each plot show the local down-sampling used to compress that spectral block. The residual between these two figures is shown in the Appendix.

In the many-featured spectral example shown above, the EELS SI files were reduced in size by more than 94% (19.4x smaller); for the 8-SI series shown in Figure 3, this corresponds to a reduction from 353MB to 18.4MB. For the Pb perovskite example shown in Figure 2, whose more simple EELS spectra contained only Sc and O edges, the compression was >54.1x.




Name	Type	Size
 EELS_Test_Spectrum.dm3	Gatan DigitalMicrograph Image...	45,270 KB
 EELS_Test_Spectrum.zip	Compressed (zipped) Folder	17,119 KB
 EELS_Test_Spectrum.lbj	LBJ File	2,325 KB

Figure 9. Example of the original and compressed EELS SI files as seen in Windows File Explorer. The compression ratio of the .lbj file is approx. 19.4x and is a further 7.4x smaller than a simple .zip file.

Similar to the EDX example presented earlier, this more than compensates for the eight frames of the MFSI acquisition, or alternatively would facilitate increasing the field-of-view significantly, or storing the final aligned spectrum where 4x digital super-resolution has been used (16x more pixels).

Discussion

As yet, there is no clear consensus in the literature as to whether total-dose or dose-rate is most responsible for electron beam damage. However, studies particularly on oxide materials present evidence that there exists some threshold dose-rate, below which damage is not observed [24,29]. In this work, we do not seek to settle this argument, but rather through the introduction of a practicable MFSI approach, we present the user with the tools to optimise their individual experiments, introducing the flexibility to fractionate dose across frames. Moreover, our first experiment with the Pb-perovskite indicates that, even when both total-dose and dose-rate are fixed, notable differences still exist based on acquisition design. Here the unit of instantaneous pixel charge is the only one which changes and further investigation of this is left to future work.

We saw from test 1, from a sample damage perspective, that it is by far preferable to record n-frames than one frame n-times slower. In addition to introducing the number of frames as an additional free-parameter for the experimentalist, data up-sampling to achieve

digital super-resolution (as used in test 2) allows for sub-pixel scanning distortions to be more precisely compensated, and for the operator to produce finely sampled chemical maps (suitable for line-profile, peak-fitting etc.) without the need to physically scan the sample at such a high pixel density. So long as the raw experimental data is recorded above the Nyquist frequency (generally $>2x$ is sufficient), then the resolvable information will be encoded in the recorded data. However, investigators generally prefer to present images far more finely sampled than this. In test 2, 4x digital up-sampling was used to improve the pixel density of the resultant chemical maps. If this density of pixels were to have been recorded experimentally, this would represent a 16x increase in experimental time and beam-exposure. In this way digital super-resolution allows the operator to optimise the acquisition to operate close to the Nyquist information limit, but to still produce finely sampled images suitable for detailed onward analysis. Moreover, recoding the experimental images at the minimum appropriate sampling is not only beneficial from a dose perspective but is also more practical for data-storage.

In test 3, we demonstrated that atomic-resolution EDX maps of repeating structures (here a single crystal), can be obtained using a sub-20pA beam more commonly associated with high-resolution imaging conditions than for analytical work. This is made possible, in part, by the extremely low dark-noise of the EDX detector and the ease of automatically aligning large multi-channel data volumes. The SNR expected in the chemical maps can be predicted using a chart such as that from Wenner et al. [8], and this is reproduced in the appendix for the material used here. As the TM approach does not use any real-space smoothing or Fourier filtering, it preserves the maximum possible resolution and does not risk introducing any 'Fourier-ringing' artefacts.

Singular value decomposition (SVD) and other blind source separation techniques are powerful statistical techniques for the reduction of noise in spectrum imaging data [30]. However, one often challenging aspect of this is determining the appropriate number of statistical components which should be used to decompose the data; too few and some important information may be missed (or under-described), while including too many components needlessly increases uncorrelated noise. The number of components is usually determined from analysing so called 'Scree plots'. However, for low SNR data, these rarely show the abrupt corner needed for an unambiguous determination and other issues may arise [31]. In this work, we observed that the separability of the orthogonal components became increasingly clear for higher doses. Gradually a second, third, and fourth component became significant rising out of the noise floor. After TM, the effective dose-accumulation yielded a sufficiently high SNR that eventually 5 components were observable; and while the material only contained four species, we attribute the need for this 5th eigenvector as being necessary to describe the two different local (de)channelling environments for the two non-equivalent silver sites. A more extensive simulation study is now underway to explore this further as a function of specimen thickness.

The data compression approaches described above for EDX and EELS not only allow for SI data to be compressed at the end of every SI-frame acquisition *before* saving to disk, but also is compatible with the serial nature of the SI distortion correction approach used in the SmartAlign algorithm [6]. In this approach, each SI can be read from disk and decompressed just as it is needed, scan-corrected, added to the running-summation and then cleared from RAM without ever needing to be written to disk in its uncompressed form. In the majority of tests performed, it was faster to compress the SI data, transmit it over a network (or write it

to a hard-drive), and decompress it than to transmit the full sized raw data. For the EDX compression, we find that the proposed compression performance actually increases as the dose-rate is reduced, offsetting the increased number of frames needed to accrue sufficient signal. For the EELS compression, while it is a prerequisite that the user must have some prior knowledge about the spectrum they will record, this is not a substantial limitation, as the user is already required to setup their spectrometer for the expected elements to be mapped, and will have likely already recorded some point-spectra in preparing the exposure conditions of their CCD. To evaluate the performance of the proposed compression further, the individual SI data shown in Figure 3 were compressed, stored, decompressed and aligned; this was compared with data which were just directly aligned with no compression. For the data where compression was used in the spectral-regions for background determination, we actually observed an improvement in the SNR of the extracted maps (see appendix). This observation is consistent with the work of Spiegelberg et al. [32] and can be attributed to a more robust extrapolation of the background in turn yielding less noisy elemental-maps.

Where the use of large fields-of-view, de-scan errors, or environmental instabilities cause energy drifts across the EELS spectrometer, these should ideally be corrected before the down-sampling compression [33,34]; this ensures the optimal size and position of the down-sampling blocks.

Future Prospects

Future possibilities for MFSI development exist in the hardware, software, and choice of investigation pursued.

Though the dark-noise of EDX spectrometers is very low, the same cannot be said of the current generation of EELS cameras. Previously, albeit for image data, we have explored systematically how increasing fractionation of a fixed dose can ultimately lead to signals that are too noisy to register [23]; we expect a similar trend to be true for EELS MFSI where increasing fractionation delivers no further benefit. This should be revisited in the future when the new generation of electron counting cameras (with greatly reduced dark-noise) become more available on spectrometers [35].

In software, the over-redundant nature of the MFSI also provides opportunities for energy-domain realignment to counter energy drift in the EELS spectrometer. This is rarely an issue in EDX, but electro-magnetic distortions, or power-supply fluctuations in the instrument can lead to *apparent* fluctuations in energy-loss [34]. The multi-frame methods described above are eminently compatible with proposed corrections for both energy-drift stability (or de-scan effects) [33,34,36], and approaches to remove systematic noise from the reference data used by stepping the spectrum across the CCD using the electrostatic drift-tube to reduce systematic artefacts that may otherwise arise from using summed spectra [14]. Where a ZLP is included in the spectrum (or with dual-EELS acquisition) the energy domain can be realigned *before* spatial alignment and spectral summing. This alignment minimises any loss of energy-resolution upon summing and allows users to achieve the fullest performance of the new generation of monochromated systems. A similar approach can also be followed to counter the energy-offset resulting from imperfect post-specimen de-scan which causes the electrons to enter the spectrometer slightly off-axis. While not the focus of this work, this is ideally suited to be incorporated in the automated acquisition and alignment scripts.

While, the ability to reduce probe-current affords operators great hope for imaging beam-sensitive materials, it also opens possibilities for studying intrinsically weak signals from beam-robust samples where currently maps are often coarsely sampled and somewhat noisy. These might include weak EELS phonon-spectroscopy [37], cathodo-luminescence (CL) [38], electron beam induced current (EBIC) [39], or electromagnetic circular di-chroism (EMCD) [40]. We hope to explore each of these opportunities in future works.

Concluding Remarks

Using multi-frame spectrum imaging we have shown that for a fixed electron budget, sample-damage can be reduced while simultaneously improving spatial precision. We have shown that with increasing electron exposure over several frames, SNR can be accumulated with no loss of spatial resolution resulting from scanning distortions; and we have shown how combining template-matching with MFSI it is possible to obtain atomic resolution EDX maps with a sub 20pA beam current. We have shown that the incorporation of digital super-resolution (pixel up-sampling *before* alignment) allows users to obtain the finely sampled chemical maps required without overly dense pixel-sampling at the point of acquisition.

To accommodate the inevitable increase in data-rate that MFSI brings, novel data-compression techniques were introduced where even for complex EELS spectra, file size reductions of 95% were achieved, while remaining locally-lossless around fine spectral details of interest. For EDX the sparsity of low-dose SI allowed for even bigger data-savings of up to 5,000x.

It is envisaged that our streamlined approach to data-acquisition, compression/storage, and non-rigid realignment will facilitate a step-change in the scope of experiment design for STEM spectroscopy.

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Author Declaration

The research described here has been partially supported by HREM Research (Tokyo) and aspects of this research will be made available as part of a commercial software plug-in for the Digital Micrograph software. A free version of the SmartAlign code (in the Matlab language) can be found at www.lewysjones.com.

References:

- [1] D. B. Williams and C. B. Carter, *Transmission Electron Microscopy*, 2nd ed. (Springer, 2009).
- [2] C. Jeanguillame and C. Colliex, *Ultramicroscopy* **28**, 252 (1989).
- [3] J. A. Hunt and D. B. Williams, *Ultramicroscopy* **38**, 47 (1991).
- [4] R. Egerton, P. Li, and M. Malac, *Micron* **35**, 399 (2004).
- [5] L. Jones and P. D. Nellist, *Microsc. Microanal.* **19**, 1050 (2013).

- [6] L. Jones, H. Yang, T. J. Pennycook, M. S. J. Marshall, S. Van Aert, N. D. Browning, M. R. Castell, and P. D. Nellist, *Adv. Struct. Chem. Imaging* **1**, 8 (2015).
- [7] B. Berkels, P. Binev, D. A. Blom, W. Dahmen, R. C. Sharpley, and T. Vogt, *Ultramicroscopy* **138**, 46 (2014).
- [8] S. Wenner, L. Jones, C. D. Marioara, and R. Holmestad, *Micron* **96**, 103 (2017).
- [9] A. B. Yankovich, C. Zhang, A. Oh, T. J. A. Slater, F. Azough, R. Freer, S. J. Haigh, R. Willett, and P. M. Voyles, *Nanotechnology* **27**, 364001 (2016).
- [10] K. Kimoto, T. Asaka, T. Nagai, M. Saito, Y. Matsui, and K. Ishizuka, *Nature* **450**, 702 (2007).
- [11] A. Stevens, L. Kovarik, H. Yang, Y. Pu, L. Carin, and N. D. Browning, *Microsc. Microanal.* **22**, 560 (2016).
- [12] J. A. Hunt and D. B. Williams, *Ultramicroscopy* **38**, 47 (1991).
- [13] F. Azough, D. M. Kepaptsoglou, Q. M. Ramasse, B. Schaffer, and R. Freer, *Chem. Mater.* **27**, 497 (2015).
- [14] M. Bosman and V. J. Keast, *Ultramicroscopy* **108**, 837 (2008).
- [15] V.-D. Hou, *Microsc. Microanal.* **15**, 226 (2009).
- [16] Y. Wang, M. R. S. Huang, U. Salzberger, K. Hahn, W. Sigle, and P. A. van Aken, *Ultramicroscopy* **184**, 98 (2018).
- [17] B. Schaffer, *How to Script... Digital Micrograph Scripting Handbook* (2017).
- [18] T. C. Lovejoy, Q. M. Ramasse, M. Falke, A. Kaeppel, R. Terborg, R. Zan, N. Dellby, and O. L. Krivanek, *Appl. Phys. Lett.* **100**, 0 (2012).
- [19] S. Hovmöller, *Ultramicroscopy* **41**, 121 (1992).
- [20] M. Haruta, Y. Fujiyoshi, T. Nemoto, A. Ishizuka, K. Ishizuka, and H. Kurata, in (2017).
- [21] F. D. La Peña, T. Ostasevicius, V. T. Fauske, P. Burdet, P. Jokubauskas, M. Nord, E. Prestat, M. Sarahan, K. E. MacArthur, D. N. Johnstone, J. Taillon, J. Caron, T. Furnival, A. Eljarrat, S. Mazzucco, V. Migunov, T. Aarholt, M. Walls, F. Winkler, B. Martineau, G. Donval, E. R. Hoglund, I. Alxneit, I. Hjorth, L. F. Zagonel, A. Garmannslund, C. Gohlke, I. Iyengar, and H.-W. Chang,

DOI:10.5281/ZENODO.583693, (2017).

- [22] S. Pack, *Factor Analysis in Chemistry*, 2nd ed. (Wiley, 1991).
- [23] L. Jones, S. Wenner, M. Nord, P. H. Ninive, O. M. Løvvik, R. Holmestad, and P. D. Nellist, *Ultramicroscopy* **179**, 57 (2017).
- [24] A. C. Johnston-Peck, J. S. DuChene, A. D. Roberts, W. D. Wei, and A. A. Herzing, *Ultramicroscopy* **170**, 1 (2016).
- [25] G. Bárcena-González, M. P. Guerrero-Lebrero, E. Guerrero, D. Fernández-Reyes, D. González, A. Mayoral, A. D. Utrilla, J. M. Ulloa, P. L. Galindo, G. Bárcena-González, M. P. Guerrero-Lebrero, E. Guerrero, D. Fernández-Reyes, D. González, A. Mayoral, A. D. Utrilla, J. M. Ulloa, and P. L. Galindo, *J. Microsc.* **0**, 1 (2015).
- [26] K. Sader, B. Schaffer, G. Vaughan, R. Brydson, A. Brown, and A. L. Bleloch, *Ultramicroscopy* **110**, 998 (2010).
- [27] S. R. Spurgeon, Y. Du, and S. A. Chambers, *Microsc. Microanal.* **1** (2017).
- [28] J. S. Jeong and K. A. Mkhoyan, *Microsc. Microanal.* **22**, 536 (2016).
- [29] N. Jiang and J. C. H. Spence, *Ultramicroscopy* **113**, 77 (2012).
- [30] N. Bonnet, N. Brun, and C. Colliex, *Ultramicroscopy* **77**, 97 (1999).
- [31] S. Lichtert and J. Verbeeck, *Ultramicroscopy* **125**, 35 (2013).
- [32] J. Spiegelberg, J. Rusz, K. Leifer, and T. Thersleff, *Ultramicroscopy* **181**, 117 (2017).
- [33] Y. Sasano and S. Muto, *J. Electron Microsc.* (Tokyo). **57**, 149 (2008).
- [34] K. Kimoto and Y. Matsui, *J. Microsc.* **208**, 224 (2002).
- [35] J. L. Hart, A. C. Lang, A. C. Leff, P. Longo, C. Trevor, R. D. Twisten, and M. L. Taheri, *Sci. Rep.* **7**, 8243 (2017).
- [36] K. Kimoto, K. Ishizuka, T. Asaka, T. Nagai, and Y. Matsui, *Micron* **36**, 465 (2005).
- [37] C. Colliex, M. Kociak, and O. Stéphan, *Ultramicroscopy* **162**, A1 (2016).
- [38] M. Kociak and O. Stéphan, *Chem. Soc. Rev.* **43**, 3865 (2014).
- [39] J. D. Poplawsky, C. Li, N. R. Paudel, W. Guo, Y. Yan, and S. J. Pennycook, *Sol. Energy Mater. Sol.*

Cells **150**, 95 (2016).

- [40] J. C. Idrobo, J. Rusz, J. Spiegelberg, M. A. McGuire, C. T. Symons, R. R. Vatsavai, C. Cantoni, and A. R. Lupini, Adv. Struct. Chem. Imaging **2**, 5 (2016).

Appendix

EDX SNR-Spacing Analysis

Following the method of Wenner et al. [8], (who operate an identical microscope to the one used for the EDX template matching in this study), the number of characteristic x-rays per square nanometre expected to be required for various projected atomic-column spacing's were evaluated.

Table 1. Calculated x-ray yields for the four species and their lateral-separation in projection.

Element	Nearest Spacing (nm)	Counts/nm ² (Single Frame)	Counts/nm ² (Σ -230 Frames)	Counts/nm ² (Template Matching)
Ag	0.3	7.5	1,725	193,150
Se	0.3	18	4,268	478,028
Sn	0.62	5	1,099	123,076
Zn	0.3	5	1,149	128,688

This experiment design tool is a descriptor which allows the operator to predict whether an experimental series will contain enough SNR even before the NR-alignment of the data has been performed.

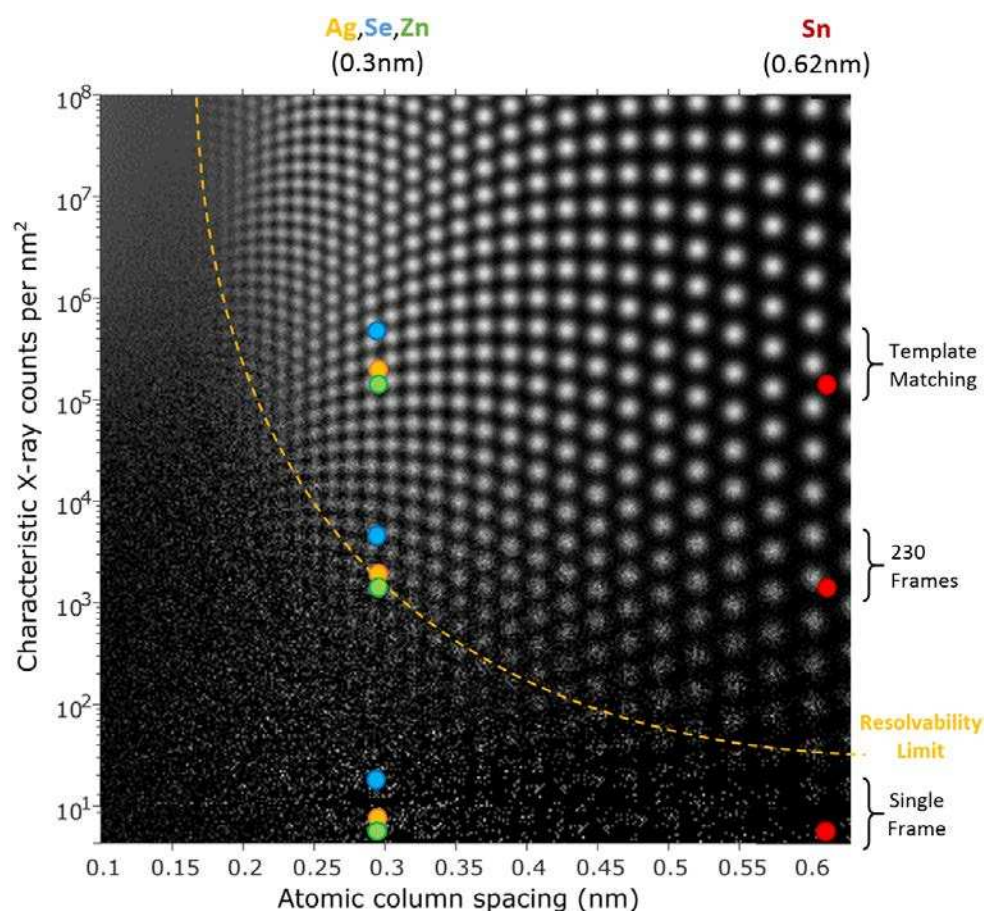


Figure A1. Simulation of the appearance of the expected EDX maps with variable characteristic x-ray signal and projected atomic-column spacing (method described in [8]). Filled circles represent the experimental points from the results discussed in the main body.

Figure A1 shows that consistent with the results shown in Figure 4, it is not expected that the single-frame EDX would be resolvable (due to the low 17pA beam current). With 230-frames of accumulated signal, the Sn map is very clearly resolved in both the predictor and the experiment, while the other elements are just marginally resolved. After template-matching (which effectively increased the x-rays per square nanometer by 112x), all four elements are clearly resolved.

EELS Compression: Further Details

The variable down-sampling used to compress the EELS has some comparisons to the 'data compression chart' method used in [2]. The equivalent chart for the method followed here is shown in Figure A2.

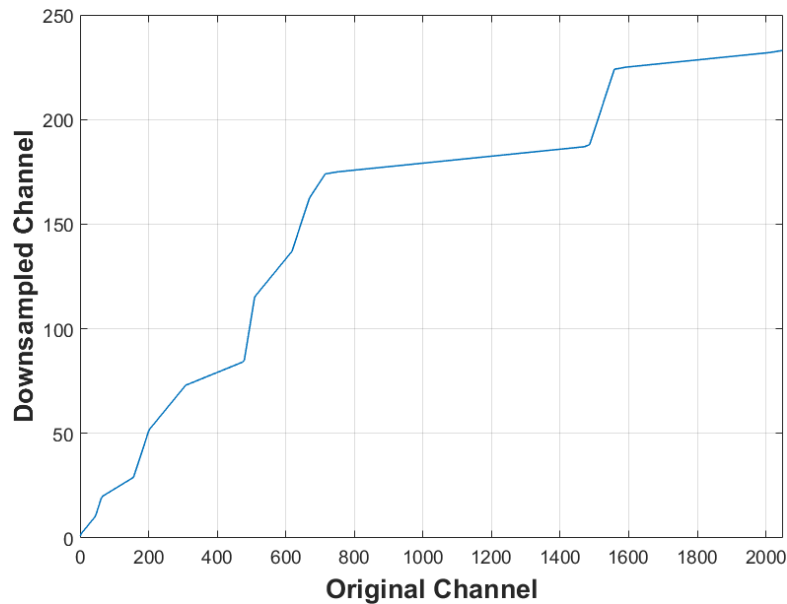


Figure A2. The equivalent 'data compression chart' representation of the down-sampling approach followed here. Shallow sections represent regions of greatest compression where primary channels are compressed into relatively fewer down-sampled points.

Taking the difference of the two spectra shown in Figure 8 yields the residual resulting from the noise-reduction, compression, and de-compression effects.

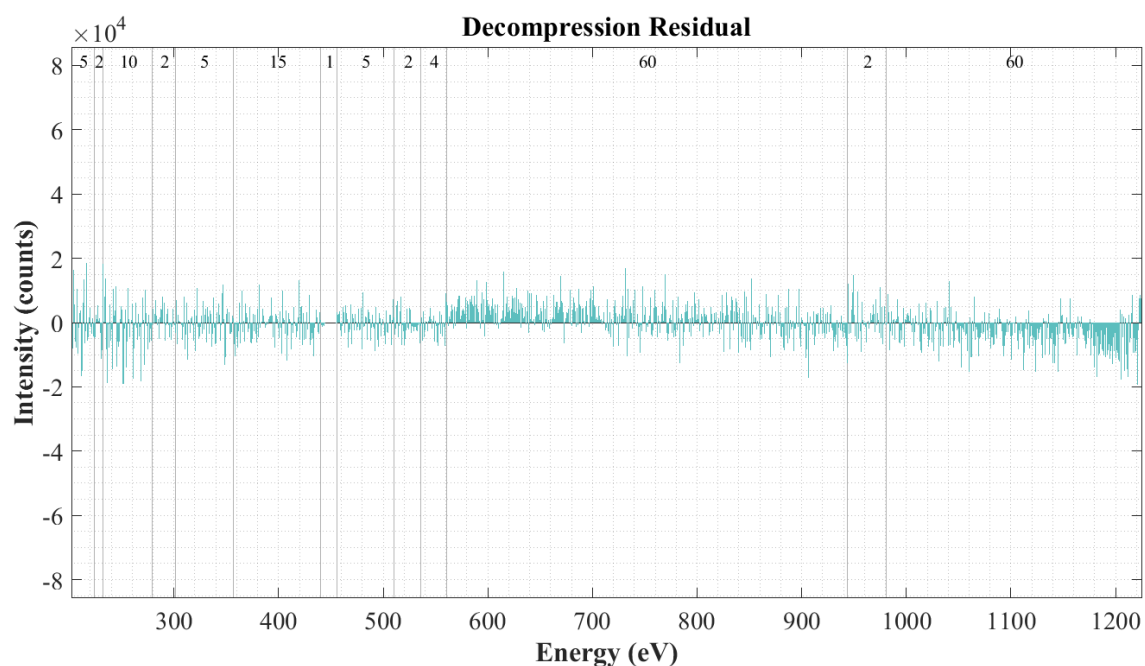


Figure A3. Residual between the raw spectrum shown in Figure 8 and its compressed and restored equivalent. The intensity scale above has been expanded a factor of ten-times larger than the figures shown in the main text

The residual is dominated by random noise apart from the uncompressed block which shows a negligibly small residual. To further compare the information-content of the restored EELS spectra (decompressed and NR-aligned), maps of the Ti (strong edge, no down-sampling) and the Nd (weak edge, 2x down-sampling) were extracted.

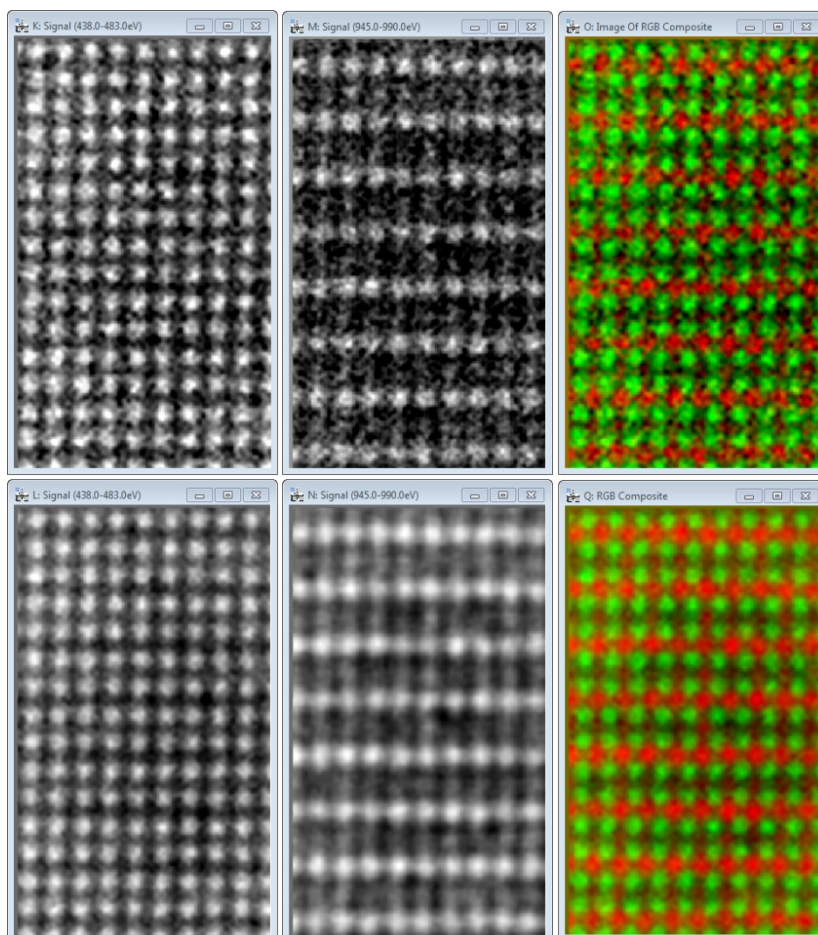


Figure A4. Extracted Ti (left) and Nd (centre) maps corresponding to the MFSI data shown in Figure 3. Top row shows the NR-aligned EELS spectra with no compression, while bottom row shows the NR-aligned spectra which were compressed and restored. The compressed and restored data appear to actually deliver a slightly improved SNR. This has been reported before and originates from the reduced noise of the pre-edge background fitting [32].